# INACTIVATED PRODUCTS OF RIFAMPICIN BY PATHOGENIC Nocardia spp.: STRUCTURES OF GLYCOSYLATED AND PHOSPHORYLATED METABOLITES OF RIFAMPICIN AND 3-FORMYLRIFAMYCIN SV

NAOKO MORISAKI and SHIGEO IWASAKI\*

Institute of Molecular and Cellular Biosciences, The University of Tokyo, 1-1-1 Yayoi, Bunkyo-ku, Tokyo 113, Japan

KATSUKIYO YAZAWA, YUZURU MIKAMI and AKIO MAEDA

Research Center for Pathogenic Fungi and Microbial Toxicoses, Chiba University, 1-8-1 Inohana, Chuo-ku, Chiba 260, Japan

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Rifampicin (1) was converted into four inactivated products by pathogenic *Nocardia*, RIP-1 and RIP-2 by *N. brasiliensis* and RIP-3 and RIP-4 by *N. otiitidiscaviarum*. MS and NMR analysis showed the compounds to be 3-formyl-23-[O-( $\beta$ -D-glucopyranosyl)]rifamycin SV (2), 23-[O-( $\beta$ -D-glucopyranosyl)]rifampicin (3), 21-(O-phosphoryl)rifampicin (4) and 3-formyl-21-(O-phosphoryl)-rifamycin SV (5), respectively.

Most pathogenic *Nocardia* are found to be resistant to rifampicin (1),<sup>1~3)</sup> a semisynthetic antibiotic widely used as a valuable chemotherapeutic agent.<sup>4)</sup> During the studies on the mechanism of the resistance,

four inactivated products of 1 were isolated, which show that the principal resistance mechanism of these organisms is the transformation of 1 to inactive compounds.<sup>4,5)</sup>

In this paper we describe the structural elucidation of the four products, RIP-1 and RIP-2 inactivated by *N. brasiliensis*, and RIP-3 and RIP-4 by *N. otitidiscaviarum*. The spectral evidence indicates the structures of RIP-1, -2, -3 and -4 as 3-formyl-23-[O-( $\beta$ -D-glucopyranosyl)]rifamycin SV (2), 23-[O-( $\beta$ -D-glucopyranosyl)]rifampicin (3), 21-(O-phosphoryl)rifampicin (4) and 3-formyl-21-(O-phosphoryl)rifamycin SV (5), respectively (Fig. 1).

#### Experimental

<sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR spectra were measured in CD<sub>3</sub>OD on a JEOL ALPHA-500 NMR spectrometer at 500, 125 and 202.35 MHz, respectively. chemical shifts of <sup>1</sup>H and <sup>13</sup>C NMR were recorded in  $\delta$  units relative to internal tetramethylsilane ( $\delta = 0$ ) and <sup>31</sup>P NMR were relative to external potassium phosphate ( $\delta = 0$ ). FAB-MS and HRFAB-MS were Fig. 1. Structures of rifampicin (1), RIP-1 (2), RIP-2 (3), RIP-3 (4) and RIP-4 (5).



 $R_2 = H, R_3 = \beta$ -D-glucose

RIP-3 (4) 
$$R_1 = CH = N - N$$
  $N - CH_3$ 

$$R_2 = -PO_3H_2, R_3 = H$$
  
RIP-4 (5)  $R_1 = CHO, R_2 = -PO_3H_2, R_3 = H$ 

Compounds	Method	Found $(m/z)$	Assignments
RIP-1	Positive FAB-MS	910	M+Na
	Negative FAB-MS	886	M-H
	HRFAB-MS	910.3518	Calcd for C <sub>44</sub> H <sub>57</sub> NO <sub>18</sub> Na
			(M + Na): 910.3474
RIP-2	Positive FAB-MS	1,007	N + Na
	Negative FAB-MS	983	M-H
	HRFAB-MS	1,007.4550	Calcd for $C_{49}H_{68}N_4O_{17}Na$
			(M + Na): 1,007.4480
RIP-3	Positive FAB-MS	903	M + H
		925	M + Na
		947	M + 2Na - H
		969	M + 3Na - 2H
	Negative FAB-MS	901	M-H
	0	923	M + Na - 2H
	HRFAB-MS	947.3471	Calcd for C <sub>43</sub> H <sub>58</sub> N <sub>4</sub> O <sub>15</sub> PNa <sub>2</sub>
			(M + 2Na - H): 947.3431
RIP-4	Positive FAB-MS	828	M + Na
		850	M + 2Na - H
		872	M + 3Na - 2H
	Negative FAB-MS	804	M-H
		826	M + Na - 2H
	HRFAB-MS	828.2602	Calcd for $C_{38}H_{48}NO_{16}PNa$ (M + Na): 828.2608

Table 1. Mass spectral data for RIPs.

Table 2. <sup>1</sup>H NMR chemical shifts ( $\delta$ , ppm) and coupling constants of rifampicin, RIP-1 and RIP-2 in CD<sub>3</sub>OD.

Proton	Rifampicin	RIP-1	RIP-2
13	1.71 (3H, s)	1.70 (3H, s)	1.69 (3H, s)
14	2.02 (3H, s)	2.02 (3H, s)	2.00 (3H, s)
17	6.35 (1H, br d, $J = 10.5 \text{ Hz}$ )	6.30 (1H, br d, $J = 11.0 \text{ Hz}$ )	6.40 (br d, $J = 10.0$ Hz)
18	7.25 (1H, dd, $J = 15.8$ , 10.5 Hz)	7.44 (1H, dd, $J = 16.0, 11.0 \mathrm{Hz}$ )	7.20 (1H, m)
19	6.08 (1H, dd, $J = 15.8$ , $7.0$ Hz)	6.14 (1H, dd, $J = 16.0, 7.5 \text{ Hz}$ )	6.04 (1H, dd, $J = 16.0, 6.5$ Hz)
20	2.31 (1H, m)	2.32 (1H, m)	2.30 (1H, m)
21	3.87 (1H, dd, $J = 10.0, 1.0$ Hz)	3.88 (1H, d, J=9.5 Hz)	3.87 (1H, d, $J = 9.0$ Hz)
22	1.74 (1H, m)	1.92 (1H, br q, $J = 7.0$ Hz)	1.83 (1H, br q, $J = 7.0$ Hz)
23	3.08 (1H, dd, J = 10.5, 2.0 Hz)	3.59 (1H, d, J=9.0 Hz)	3.56 (1H, d, J=9.0  Hz)
24	1.48 (1H, m)	1.54 (1H, m)	1.56 (1H, m)
25	5.16 (1H, d, $J = 10.5$ Hz)	5.39 (1H, d, $J = 10.5$ Hz)	5.34 (1H, d, $J = 10.5$ Hz)
26	1.24 (1H, m)	1.02 (1H, m)	1.01 (1H, m)
27	3.38 (1H, d, J = 8.0 Hz)	3.34 (1H, d, J = 8.5 Hz)	3.34 (1H, dd, J = 8.5, 1.5 Hz)
28	5.07 (1H, dd, J = 12.7, 8.0 Hz)	5.09 (1H, dd, $J = 12.5$ , 8.5 Hz)	5.14 (1H, dd, J = 12.5, 8.5 Hz)
29	6.26 (1H, d, $J = 12.7$ Hz)	6.31 (1H, d, $J = 12.5$ Hz)	6.27 (1H, d, J = 12.5 Hz)
30	2.02 (3H, s)	1.98 (3H, s)	2.02 (3H, s)
31	0.93 (3H, d, $J = 7.0$ Hz)	1.01 (3H, d, $J = 7.0$ Hz)	0.94 (3H, d, J = 7.0 Hz)
32	0.99 (3H, d, J = 7.0 Hz)	1.08 (3H, d, $J = 7.0$ Hz)	1.03 (3H, d, $J = 7.0$ Hz)
33	0.61 (3H, d, $J = 7.0$ Hz)	0.60 (3H, d, J = 7.0 Hz)	0.54 (3H, d, J = 7.0 Hz)
34	-0.21 (3H, d, $J = 7.0$ Hz)	-0.11 (3H, d, $J = 7.0$ Hz)	0.03 (3H, d, J = 7.0 Hz)
36	2.02 (3H, s)	2.02 (3H, s)	2.02 (3H, s)
37	3.00 (3H, s)	3.01 (3H, s)	3.01 (3H, s)
$N-CH_3$	2.78 (3H, s)	—	2.42 (3H, s)
PhCH=N-N	8.32 (1H, s)	·i	8.19 (1H, s)
PhCH=O		10.52 (1H, s)	
$CH_2N$	3.30 (4H, br m) <sup>a</sup> ,		3.18 (4H, brm),
	3.18 (4H, br m)		2.72 (4H, br m)
Glu-1'		4.42 (1H, d, $J = 7.5$ Hz)	4.40 (1H, d, $J = 7.8$ Hz)
Glu-2'		2.97 (1H, dd, $J=9.5$ , 7.5 Hz)	2.99 (1H, dd, $J = 9.0$ , 7.8 Hz)
Glu-3'		3.28 (1H, dd, J=9.5, 9.0 Hz)	3.26 (1H, dd, J=9.5, 9.0 Hz)
Glu-4'		3.15 (1H, dd, J=9.0, 8.0 Hz)	3.16 <sup>a</sup>
Glu-5'		3.17 (1H, ddd, J=8.0, 4.5, 1.5 Hz)	3.16 <sup>a</sup>
Glu-6'		3.58 (1H, dd, J = 12.5, 4.5 Hz),	3.59 (1H, dd, J = 12.5, 5.0 Hz),
		3.81 (1H, dd, J = 12.5, 1.5 Hz)	3.81 (1H, dd, J = 12.5, 1.5 Hz)

<sup>a</sup> Obscured by other resonances.

Carbon	Rifampicin	RIP-1	RIP-2	Carbon	Rifampicin	RIP-1	RIP-2
$1 \sim 10$	184.0	185.3	185.6	25	75.6	77.3	77.3
	175.8	174.2	174.7	26	41.7	42.8	42.8
	149.3	152.3	149.0	27	78.5	78.7	78.9
	147.9	148.8	147.1	28	120.1 <sup>b</sup>	119.9	120.0
	119.7 <sup>b</sup>	120.3	118.8	29	144.7	145.4	145.1
	118.2	120.2	117.9	30	20.8	20.3	20.7
	116.1	119.2	116.9	31	18.2	18.3	18.4
	116.0	115.3	115.9	32	11.0	11.9	12.3
	105.0	105.7	104.3	33	9.4	10.9	10.9
	101.9	101.7	101.3	34	9.7	9.5	9.8
11	189.0	188.6	187.3	35	172.4	172.9	172.9
12	110.6	110.6	110.5	36	20.8	21.0	21.0
13	22.4	22.4	22.3	37	56.7	56.4	56.4
14	7.5	7.4	7.4	NCH <sub>3</sub>	43.7		45.4
15	171.1	171.6	174.7	PhCH = N-N	138.8		136.8
16	133.1	133.3	133.2	PhCH=O	_	193.1	
17	134.8	134.1	135.1	$CH_2N$	53.2	_	54.6
18	129.0	129.7	128.6		49.5ª		50.9
19	140.7	140.0	141.0	Glu-1'		104.0	103.9
20	39.1	39.3	40.3	Glu-2'		76.0	75.9
21	75.2	76.0	74.0	Glu-3'		78.1°	78.1 <sup>d</sup>
22	34.4	36.0	36.4	Glu-4'		71.3	71.4
23	78.2	88.9	88.5	Glu-5'		78.2°	78.2 <sup>d</sup>
24	39.4	39.6	39.5	Glu-6'		62.7	62.8

Table 3. <sup>13</sup>C NMR chemical shifts ( $\delta$ , ppm) of rifampicin, RIP-1 and RIP-2 in CD<sub>3</sub>OD.

<sup>a</sup> Obscured by other resonances.

<sup>b~d</sup> Interchangeable.

measured on a JEOL HX110 instrument. Biological transformation of rifampicin to RIPs, the isolation and purification are described elsewhere.<sup>4,5)</sup>

## Structure

The structures of **RIPs** were determined spectroscopically by comparison with the data of rifampicin.

Positive FAB-MS of RIP-1 showed a peak at

m/z 910 (M + Na)<sup>+</sup>, and negative FAB-MS at m/z 886 (M – H)<sup>-</sup>. The molecular formula was determined by HRFAB-MS to be C<sub>44</sub>H<sub>57</sub>NO<sub>18</sub> (Table 1).

The structure of RIP-1 was elucidated to be 3-formyl-23-[O-( $\beta$ -D-glucopyranosyl)]rifamycin SV (2) by <sup>1</sup>H NMR, <sup>13</sup>C NMR, COSY, C-H COSY and HMBC experiments. <sup>1</sup>H NMR spectrum of RIP-1 (Table 2) showed a singlet due to a formyl proton at  $\delta$  10.52 instead of the signals of *N*-methyl piperazine moiety ( $\delta$  2.78, 3.30, 3.18) and of an olefinic proton on the carbon adjacent to C-3 ( $\delta$  8.32) present in rifampicin,<sup>6</sup> indicating the 1-amino-4-methyl piperazine moiety has been cleaved off by hydrolysis. The formyl carbon signal was also shown at  $\delta$  193.1 in <sup>13</sup>C NMR spectrum (Table 3). Protons 17-H through 21-H and 25-H through 29-H were correlated by COSY spectrum and carbon signals of C-15 through C-29 were assigned by HMBC experiment. The presence of D-glucose was shown by signals at  $\delta$  104.0 (C-1'), 76.0 (C-2'), 78.1 (C-3'), 71.3 (C-4'), 78.2 (C-5') and 62.7 (C-6') in the <sup>13</sup>C NMR spectrum.<sup>7</sup> The site of glycosylation was determined by HMBC experiment (Fig. 2) to correlate 23-H ( $\delta$  3.59) with C-1', and 1'-H ( $\delta$  4.42) with C-23 ( $\delta$  88.9). Down field shifts of the 23-H signal ( $\Delta\delta$  0.51 ppm) and C-23 signal ( $\Delta\delta$  10.7 ppm) relative to those of rifampicin also indicated the glycosylation of the OH group at C-23. The coupling constant of the anomeric proton at  $\delta$  4.42 (d, J=7.5 Hz) is consistent with the  $\beta$ -glucoside. No other significant differences were observed in the NMR and UV spectra of RIP-1 and rifampicin, indicating that the rest of the structures of these two compounds are the same.





Proton	RIP-3	RIP-4
13	1.69 (3H, s)	1.71 (3H, s)
14	1.99 (3H, s)	$1.99 (3H, s)^d$
17	6.33 (1H, br m)	6.30 (1H, d, $J = 11.0$ Hz)
18	6.98 (1H, br m)	7.16 (1H, br m)
19	5.83 (1H, br dd, $J = 15.0, 9.0 \text{ Hz}$ )	5.92 (1H, dd, $J = 15.0, 8.0 \text{Hz}$ )
20	2.22 (1H, br m)	2.30 (1H, br m)
21	4.28 (1H, br m)	4.23 (1H, br m)
22	1.81 (1H, br m)	1.88 (1H, br m)
23	2.81 (1H, dd, $J=9.5$ , 3.0 Hz)	2.84 (1H, dd, $J = 10.0$ , 2.5 Hz)
24	1.43 (1H, br m)	1.52 (1H, m)
25	5.01 (1H, br d, $J=9.5$ Hz)	5.08 (1H, d, $J = 11.0$ Hz)
26	1.0 <sup>b</sup> (1H, br m)	0.93 (1H, br m)
27	3.3 <sup>b</sup>	3.3 <sup>b</sup>
28	5.25 (1H, dd, J=12.5, 8.0 Hz)	5.27 (1H, dd, $J = 12.5$ , 7.5 Hz)
29	6.16 (1H, d, $J = 12.5$ Hz)	6.19 (1H, d, $J = 12.5$ Hz)
30	1.97 (3H, s) <sup>c</sup>	2.02 (3H, s) <sup>d</sup>
31	1.13 (3H, d, J = 7.0 Hz)	1.12 (3H, d, J = 7.0 Hz)
32	0.91 (3H, d, J = 7.0 Hz)	0.97 (3H, d, J = 7.0 Hz)
33	0.21 (3H, d, $J = 7.0$ Hz)	$0.22 (3H, d, J = 7.0 Hz)^{e}$
34	0.31 (3H, br m)	0.23 (3H, d, $J = 7.0 \text{ Hz})^{e}$
36	2.01 (3H, s)°	2.02 (3H, s) <sup>d</sup>
37	3.02 (3H, s)	3.02 (3H, s)
NCH <sub>3</sub>	2.49 (3H, s)	
PhCH=N-N	8.17 (1H, s)	
PhCH=O		10.50 (1H, s)
$CH_2N$	3.18 (4H, br m), 2.71 (4H, br m)	

Table 4. <sup>1</sup>H NMR chemical shifts ( $\delta$ , ppm) and coupling constants of RIP-3 and RIP-4 in CD<sub>3</sub>OD<sup>a</sup>.

<sup>a</sup> The data of rifampicin is shown in Table 2.

<sup>b</sup> Obscured by other resonances.

°~° Interchangeable.

The structure of RIP-2 was elucidated to be 23-[O-( $\beta$ -D-glucopyranosyl)]rifampicin (3) by the same procedures as used for RIP-1. Positive FAB-MS showed a peak at m/z 1,007 (M+Na)<sup>+</sup>, and negative FAB-MS at m/z 983 (M-H)<sup>-</sup>. Its molecular formula was determined by HRFAB-MS to be C<sub>49</sub>H<sub>68</sub>N<sub>4</sub>O<sub>17</sub> (Table 1) which is in accord with a monoglucosylated rifampicin, and this was confirmed by its <sup>1</sup>H NMR and <sup>13</sup>C NMR data (Tables 2 and 3). The site of glycosylation was similarly determined by HMBC spectrum to correlate 23-H ( $\delta$  3.56) with C-1' ( $\delta$  103.9) and 1'-H ( $\delta$  4.40) with C-23 ( $\delta$  88.5). Both the 23-H signal and the C-23 signal shifted to downfield ( $\Delta\delta$  0.48 ppm and  $\Delta\delta$  10.3 ppm, respectively) compared

Fig. 3. <sup>1</sup>H-<sup>1</sup>H correlation of RIP-3 (4) by COSY experiment.



with those signals of rifampicin. Thus, the structure of RIP-2 was established to be  $23-[O-(\beta-D-glucopyranosyl)]$ rifampicin (3).

Positive and negative FAB-MS data of RIP-3 and RIP-4 indicated their molecular weight to be 902 and 805, respectively. The molecular formulae were determined by HRFAB-MS to be  $C_{43}H_{59}N_4O_{15}P$  for RIP-3 and  $C_{38}H_{48}NO_{16}P$  for RIP-4 (Table 1). <sup>31</sup>P NMR spectrum of RIP-3 and of RIP-4 showed a signal at  $\delta$  2.05 and  $\delta$  1.79, respectively, due to a phosphoric acid ester. These data suggest that RIP-3 and RIP-4 are *O*-phosphorylated rifampicin and phosphorylated 3-formylrifamycin SV, respectively, instead

Fig. 4. COSY spectrum of RIP-4 (5).



of the glycosylation in the case of RIP-1 and RIP-2.

The structure of RIP-3 was determined by <sup>1</sup>H NMR (Table 4), COSY (Fig. 3) and <sup>31</sup>P NMR experiments. In COSY experiment, <sup>1</sup>H signals of 17-H through 24-H and of 25-H through 29-H were correlated, and the signals at  $\delta$  4.28 and at  $\delta$  2.81 were assigned to 21-H and 23-H, respectively. Comparing the chemical shifts of <sup>1</sup>H-signals with those of rifampicin (Table 2), prominent down field shift caused by phosphorylation was observed only for 21-H ( $\Delta\delta$  0.41 ppm).<sup>8,9</sup> In a {H}-P selective decoupling experiment, a doublet at  $\delta$  2.05 ( $J_{21-H,P}$ =8.4 Hz)<sup>10</sup> due to <sup>31</sup>P callapsed into a sharp singlet by irradiation at  $\delta$  4.28 (21-H). These facts indicate that 21-OH of rifampicin was phosphorylated in RIP-3 (4).

<sup>1</sup>H NMR spectrum of RIP-4 lacked the signals of *N*-methyl piperazine moiety present in rifampicin and, instead, exhibited a signal due to a formyl proton at  $\delta$  10.50 (Table 4). The signals of 21-H ( $\delta$  4.23) and 23-H ( $\delta$  2.84) were assigned based on the correlation of 19-H through 24-H in COSY experiments (Fig. 4). Down field shift of 21-H ( $\Delta$  0.36 ppm) indicates that phosphorylation occured, like RIP-3, on the 21-OH.

No other significant differences was observed in the <sup>1</sup>H NMR spectra of RIP-3, RIP-4 and rifampicin, indicating that the rest of the structures of these compounds are the same. Thus, the structures of RIP-3 and RIP-4 were determined to be 21-(O-phosphoryl)rifampicin (4) and 3-formyl-21-(O-phosphoryl)-rifamycin SV (5), respectively.

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